

Light sources and conditions for photoactivation of aryl azide crosslinking and labeling reagents

TR0011.1

Introduction

Pierce offers a variety of photoreactive crosslinking and biotinylation reagents (see Related Pierce Products). These are useful for many applications including the study of protein:protein interactions, isolating cell surface proteins and preparing labeled probes. With few exceptions, the photoreactive groups in these reagents are aryl azides (Figure 1). When an aryl azide is exposed to UV-light, it forms a nitrene group that can initiate addition reactions with double bonds, insertion into C-H and N-H sites, or subsequent ring expansion to react as a nucleophile with primary amines (Figure 2). The latter reaction path dominates when primary amines are present in the sample.

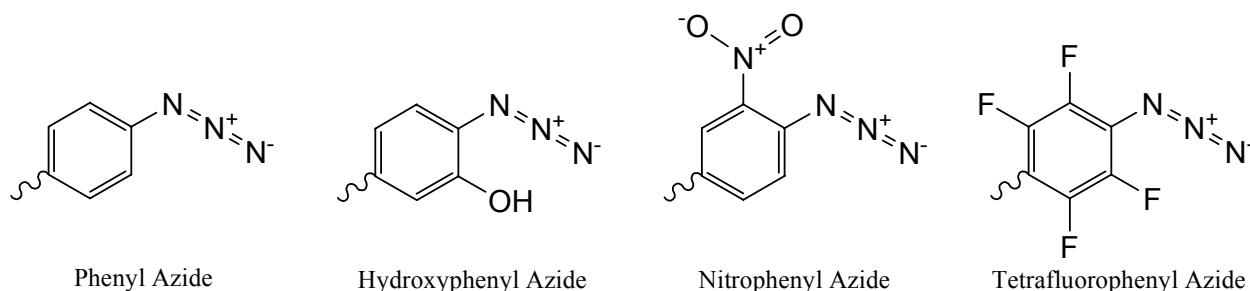


Figure 1. Forms of aryl azide reactive groups in photoreactive crosslinking reagents.

A question that often arises with respect to aryl azide linkers is what wavelength and intensity of light is optimal for photoactivation and efficient crosslinking. Pierce researchers have performed only a limited number of experiments to explore these conditions. However, one set of data was generated for the homobifunctional hydroxyphenyl azide crosslinker BASED (Product No. 21564). Experiments with three different lamp sources and several exposure times indicated that activation with long wavelength UV-light (366 nm) for 30 minutes yields the most efficient (complete) crosslinking of the target molecule (in this case, a peptide) and depletion of free crosslinker (Table 1). These data for BASED are probably representative for all the aryl azide reagents, although short wavelength may be better than long wavelength for plain phenyl azides (no hydroxy or nitro group on the phenyl ring). Examination of Table 1 and perusal of the literature (Table 2) indicate that successful photoactivation with these reagents is possible across a wide range of wavelength and time of exposure.

In addition to choosing an appropriate lamp source for photoactivation, consider the following points when preparing conjugation reactions with aryl azide crosslinkers:

- When microcentrifuge tubes are used for the sample, it is most effective to open the cap so that the sample may be exposed directly rather than through the polypropylene sidewall, which shields most of the UV-light. Quartz spectrophotometric cuvettes are an ideal choice for reaction vessels because they allow for optimal exposure of the sample to the UV-light source.
- Samples will become warm or even hot if exposed to intense UV-light for several minutes. Place the sample vial on ice or use some other method to keep the sample cool during UV-light activation.
- Avoid thiol-containing reducing agents (e.g., DTT or 2-mercaptoethanol) in the sample solution during all steps before and during photoactivation. These reagents will reduce the azide functional group to an amine, preventing photoactivation.
- Avoid buffers that contain primary amines (e.g., Tris or glycine) during photoactivation because these will quench the desired reaction. Reaction of the photoactivated aryl azide groups to primary amines dominates if they are present.

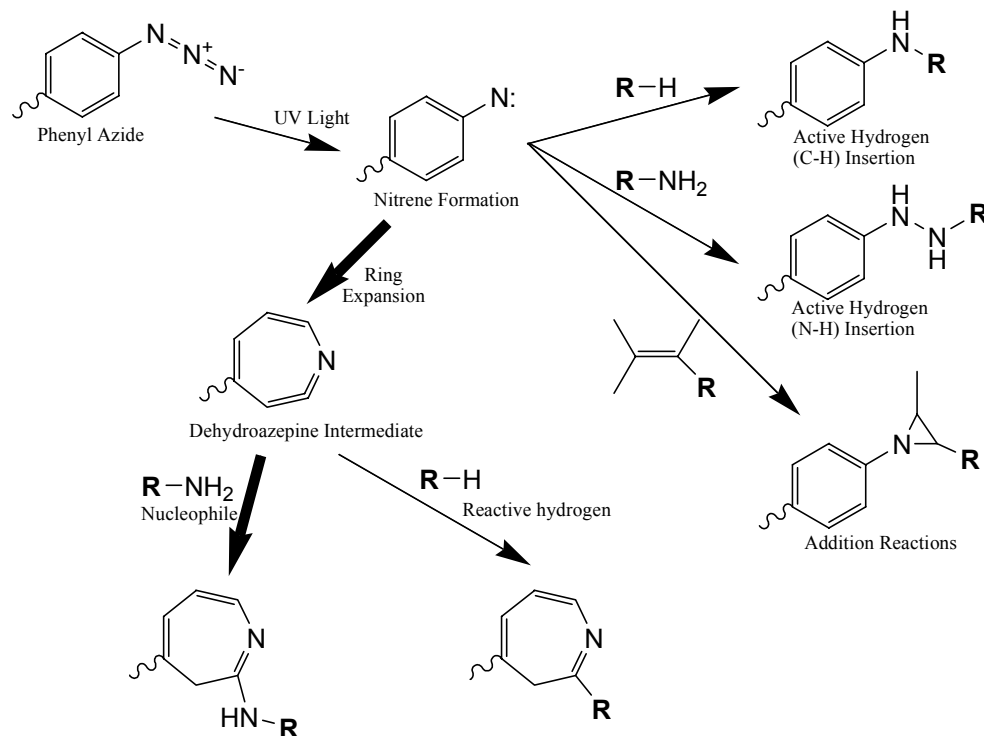


Figure 2. Possible reaction pathways of aryl azide crosslinkers.

Table 1. Lamp conditions and crosslinking efficiency using BASED (Product No. 21564). Identical reactions with BASED and peptide were prepared and photoactivated by exposure at different wavelengths and lengths of time. Conjugation efficiency was determined by HPLC measurement of the percent peptide and crosslinker depleted relative to the starting material (greater depletion corresponds to more complete crosslinking).

Sample	Time	% Peptide depleted	% BASED depleted
Long Wave UV-Light (366 nm)	5 min	41.46	46.79
	15 min	47.92	77.14
	30 min	61.46	94.33
Short Wave UV-Light (254 nm)	5 min	11.26	9.61
	15 min	—	—
	30 min	14.46	24.82
550 Watt Light (Broad spectrum visible)	10 sec	27.87	31.51
	30 sec	—	30.52
	60 sec	42.43	58.54
550 Watt Light (Broad spectrum visible)	6 flashes, each 1 sec	3.82	18.38
	12 flashes, each 1 sec	19.50	29.95
	18 flashes, each 1 sec	12.26	32.23
Combination	15 min Long UV-Light	35.23	74.60
	Plus 12 flashes	37.76	76.54

Table 2. Lamp sources and conditions used for photoactivation of aryl azide crosslinkers. The information in this table was compiled from older (pre-2000) literature references and is provided only to exemplify the types of conditions used by researchers.

Lamp	Distance from sample	Wavelength	Time
Rayonet Photochemical Reactor (UltraViolet Products, Inc.)	2 cm	300 nm	?*
Model UVGL-15 Mineralight	?	254-360 nm	10 min
?	3.4 cm	366 nm	5 min
Fotodyne UV Transilluminator	5 cm	?	2 min
750 W mercury lamp (Scaeffel Instrument Co.)	20 cm	320 nm	?
Black Ray model XX-15C, 0.41 A (Black Ray UV Products, Inc.)	18 cm 2.5 cm 10 cm	? ? ?	75 min 2-30 min 5 min
Edmund Scientific No. 60889 (9 W, 12 in. UV tube)	2 mm	>300 nm	120 min
Mineralight UVS-11	1 cm	?	5 min
Black Ray Model B-100A	10 cm	>300 nm	5 min
ACME LITE model 228A electronic flash (cover removed)	1 cm	?	20 flashes
Transilluminator UV light box (UltraViolet Products, Inc.)	3.5 cm	302 nm	5 min
Sun Thyristor Flash unit (1/4 power)	2 cm	?	20 flashes
500 W Xenon lamp	9 cm	?	2-10 min
Zeiss 100 W mercury lamp	15 cm	?	?
250 W sunlamp (General Electric)	15 cm	?	15 min
15 W UV lamp	1 cm	365 nm	10 min
Kako Auto 201 electronic flash (plastic cover removed)	3-5 cm	?	5 flashes
50 W HBO mercury lamp (OSRAM, Berlin)	?	?	30 sec
UVS-11 Mineralight	5 cm	?	3 min
100 W HBO-101 xenon lamp (Narva, Berlin)	1 cm	?	70 sec
UVS-52	15 cm	?	3 min
Rayonet UV Light Reactor (Southern N.E. Ultraviolet Co.)	10 cm	370 nm	5 min
500 W xenon lamp	9 cm	?	10 sec
Chromato-Vue C3 viewing box (Ultraviolet Products, Inc.)	10 cm	254 and 365 nm	20 min
Model AH-6 1 kW high pressure Hg lamp (Advanced Radiation)	10 cm	?	5 sec
Universal UV lamp (Camag Muttentz, Switzerland)	1 cm	254 nm	2 min
UV SL-25 4 W Mineralight (Ultraviolet Products, Inc.)	1 cm	254 nm	10 min

Related Pierce Products (Photoreactive reagents offered by Pierce)

Product Name	Product No.	Reactive Groups		Spacer Arm	
		Photoreactive	Other Group(s)	Length (Å)	Cleavable?
ABH	21510	Phenyl azide	Hydrazide	11.9	No
ANB-NOS	21451	Nitrophenyl azide	NHS	7.7	No
APDP	27720	Hydroxyphenyl azide	Pyridyldisulfide	21.0	Yes
APG	20108	Phenyl azide	Phenylglyoxal	9.3	No
ASBA	21512	Hydroxyphenyl azide	Amine	16.3	No
BASED	21564	Hydroxyphenyl azide	Hydroxyphenyl azide	34.7	Yes
Mts-Atf-Biotin Mts-Atf-LC-Biotin	33093 33083	Tetrafluorophenyl azide	Methanethiosulfonate and Biotin	11-31 21-35	Yes
NHS-ASA	27714	Hydroxyphenyl azide	NHS	8.0	No
SADP	21533	Phenyl azide	NHS	13.9	Yes
SANPAH	22600	Nitrophenyl azide	NHS	18.2	No
SPB	23019	Psoralen	NHS	8.6-9.5	No
Sulfo-HSAB	21563	Phenyl azide	Sulfo-NHS	9.0	No
Sulfo-NHS-LC-ASA	27735	Hydroxyphenyl azide	Sulfo-NHS	18.0	No
Sulfo-SADP	21553	Phenyl azide	Sulfo-NHS	13.9	Yes
Sulfo-SAED	33030	Azido-methylcoumarin	Sulfo-NHS	23.6	Yes
Sulfo-SAND	21549	Nitrophenyl azide	Sulfo-NHS	18.5	Yes
Sulfo-SASD	27716	Hydroxyphenyl azide	Sulfo-NHS	18.9	Yes
Sulfo-SFAD	27719	Perfluoroaryl azide	Sulfo-NHS	14.6	Yes
Sulfo-SANPAH	22589	Nitrophenyl azide	Sulfo-NHS	18.2	No
Sulfo-SBED	33033	Phenyl azide	Sulfo-NHS/Biotin	14-25	Yes
Psoralen-PEO-Biotin	29986	Psoralen	Biotin	36.9	Yes
Photoactivatable Biotin	29987	Nitrophenyl azide	Biotin	30.0	No
Biotin-LC-ASA	29982	Hydroxyphenyl azide	Biotin	29.9	No
TFPA-PEO₃-Biotin	21303	Tetrafluorophenyl azide	Biotin	33.4	No

References

- Hermanson, G. T. (1996). Bioconjugate Techniques, Academic Press, Inc., p. 163, 252-255.

Note: This book is available from Pierce as Product No. 20002.

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